

101.121-24/04 – including *Taq* pol., IFU-01
101.121-24u/04u – without *Taq* pol., IFU-02

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“Instructions for Use” (IFU)

Lot No.: **34X**

Lot-specific information
Olerup SSP® DRB3

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Product number:	101.121-24/04 – including <i>Taq</i> pol. 101.121-24u/04u – without <i>Taq</i> pol.
Lot number:	34X
Expiry date:	2017-April-01
Number of tests:	24 tests – Product No. 101.121-24/24u 4 tests – Product No. 101.121-04/04u
Number of wells per test:	31+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 34X.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP®
DRB3 Lot (94S)**

A well containing Negative Control primer pairs has been added.

The format of the Product Insert and Worksheet have been changed.

The DRB3 primer set as well as the specificity and interpretation tables have been updated for the DRB alleles described since the previous *Olerup SSP®* DRB3 lot (**Lot No. 94S**) was made. The kit design is based on IMGT/HLA database 3.17.0.

As of lot series V, the Specificity Table is included in the lot-specific Product Insert, and the Interpretation Table is included in the Worksheet.

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The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
4	Moved	Removed	5'-primer moved to wells 14, 22 and 23. 3'-primer removed.
14	Added	-	5'-primer added from well 4.
22	Added	-	5'-primer added from well 4.
23	Added	-	5'-primer added from well 4.
32	Moved	Moved	Primer pair moved to well 4, negative control.

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Well **32** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP®* HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80	85
5'-primer¹	164	340	440	45	45	43	36
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-TAC ^{3'}
							36
							5'-TAT ^{3'}
3'-primer²	231	2nd I	507	59	58	57	47
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}	5'-ACA ^{3'}
							48
							5'-gCA ^{3'}
							48
							5'-gCC ^{3'}
							52
							5'-TgT ^{3'}
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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PRODUCT DESCRIPTION

DRB3 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DRB3*01:01 to DRB3*03:03 alleles.

PLATE LAYOUT

Each test consists of 32 PCR reactions in a 32 well cut PCR plate.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	16
17	18	19	20	21	22	23	24
25	26	27	28	29	30	31	NC

The 32 well cut PCR plate is marked with 'DRB3' in silver/gray ink.

Well No. 1 is marked with the Lot No. '34X'.

Wells 1 to 31 – DRB3 high resolution primers.

Well 32 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 32 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between DRB3 and DRB1 alleles, primer mixes 1 to 5, 7 to 14, 16, 17, 19, 20, 24, 26, 28, 29 and 31 will amplify DRB1 alleles.

For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the DRB3 alleles, i.e. **DRB3*01:01 to DRB3*01:15, DRB3*02:01 to DRB3*02:29N and DRB3*03:01 to DRB3*03:03**, recognized by the HLA Nomenclature Committee in July 2014^{1,2} will give rise to unique amplification patterns by the primers in the DRB3 subtyping kit.

The DRB3 subtyping kit cannot distinguish the silent mutations in the DRB3*01:01:02:01 to 01:01:05 alleles, the DRB3*02:02:01:01 to 02:02:05 alleles or the DRB3*03:01:01 to 03:01:03 alleles.

¹DRB alleles listed on the IMGT/HLA web page 2014-July-25, release 3.17.0, www.ebi.ac.uk/imgt/hla.

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²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page
<http://hla.alleles.org/alleles/deleted.html>.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in DRB3 homo- and heterozygotes is available upon request.

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SPECIFICITY TABLE

DRB3 SSP subtyping

Specificities and sizes of the PCR products of the 31+1 primer mixes used for DRB3 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DRB3 alleles ³	Other amplified DRB alleles ⁴
1⁵	100 bp	515 bp	*01:01:02:01-01:08, 01:10-01:13, 01:15, 02:05, 03:01-01-03:01:03, 03:03	DRB1*03:42, DRB1*03:87, DRB1*11:30 ^w , DRB1*13:67, DRB1*14:46
2⁵	125 bp	430 bp	*01:01:02:01-01:06, 01:08, 01:10-01:14, 03:01-01-03:01:03, 03:03	DRB1*03:38, DRB1*03:79, DRB1*03:85, DRB1*03:96, DRB1*04:82 ^w , DRB1*04:99 ^w , DRB1*08:07 ^w , DRB1*08:19 ^w , DRB1*08:25 ^w , DRB1*08:34 ^w , DRB1*08:52 ^w , DRB1*12:27 ^w , DRB1*13:31 ^w , DRB1*13:46 ^w , DRB1*13:54 ^w , DRB1*13:77 ^w , DRB1*13:100 ^w , DRB1*13:162 ^w , DRB1*13:180 ^w , DRB1*14:48 ^w
3⁵	95 bp	430 bp	*01:01:02:01-01:02, 01:04-01:08, 01:10-01:13, 01:15	DRB1*03:42
4⁸	240 bp	430 bp		DRB1*11:13:01-11:13:02, DRB1*11:149, DRB1*13:08, DRB1*14:01:01, DRB1*14:01:02 [?] -14:01:04 [?] , DRB1*14:02:02 [?] , DRB1*14:04-DRB1*14:05:01, DRB1*14:05:02 [?] -14:05:04 [?] , DRB1*14:06:02 [?] -14:06:03 [?] , DRB1*14:07:01, DRB1*14:07:02 [?] , DRB1*14:08, DRB1*14:09 [?] -14:10 [?] , DRB1*14:11, DRB1*14:12:01 [?] -DRB1*14:12:02 [?] , DRB1*14:14, DRB1*14:15 [?] -14:20 [?] , DRB1*14:22 [?] -DRB1*14:45 [?] , DRB1*14:47 [?] -14:53 [?] , DRB1*14:54:01-14:54:04, DRB1*14:55 [?] -14:56 [?] , DRB1*14:57, DRB1*14:58 [?] -14:65 [?] , DRB1*14:67 [?] -DRB1*14:81 [?] , DRB1*14:82, DRB1*14:83 [?] -DRB1*14:136 [?] , DRB1*14:137N, DRB1*14:138 [?] -14:140 [?] , DRB1*14:141, DRB1*14:142 [?] -14:143 [?] , DRB1*14:145 [?] -DRB1*14:151 [?] , DRB1*14:152N, DRB1*14:153 [?] -14:157 [?]
5⁵	95 bp 125 bp	430 bp	*01:03 *01:10	DRB1*14:46
6	190 bp	430 bp	*01:04	
7^{5,7}	90 bp 180 bp	515 bp	*01:05 *01:07, 01:15	DRB1*03:42, DRB1*14:46
8⁵	120 bp	430 bp	*01:06, 01:08	DRB1*03:42, DRB1*14:46

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9⁶	165 bp	430 bp	*01:07, 02:01-02:08, 02:11-02:29N	DRB1*14:141
10⁶	185 bp	515 bp	*01:09, 02:01-02:04, 02:06-02:14, 02:15 ^w , 02:16-02:26, 02:27 ^w , 02:28-02:29N, 03:02	DRB1*03:101, DRB1*08:32^w, DRB1*12:01:02^w-12:01:03^w, DRB1*12:02:02^w, DRB1*12:03:02^w, DRB1*12:16:01^w, DRB1*12:16:03^w, DRB1*12:22^w, DRB1*14:121^w, DRB1*14:141, DRB1*15:03:01:01^w-15:03:02^w, DRB1*15:23^w, DRB1*15:54^w, DRB1*15:57^w, DRB1*15:78^w, DRB1*15:94^w
11⁷	270 bp	515 bp	*02:01, 02:04, 02:24, 03:01:01-03:02	DRB1*03:42, DRB1*03:87
12⁵	90 bp 145 bp	515 bp	*02:01, 02:04, 02:17, 02:24	DRB1*03:37, DRB1*03:80, DRB1*11:43, DRB1*11:50, DRB1*11:115, DRB1*11:151, DRB1*12:34, DRB1*13:131, DRB1*13:150, DRB1*14:59, DRB1*14:86, DRB1*14:96, DRB1*14:145, DRB1*14:153-14:154, DRB1*15:65
13⁶	270 bp	515 bp	*01:01:02:01-01:13, 01:15, 02:02:01:01-02:03, 02:05-02:13, 02:15-02:23, 02:25-02:29N, 03:03	DRB1*11:30, DRB1*13:67, DRB1*14:46, DRB1*14:141
14	185 bp	515 bp	*01:07, 01:09, 02:02:01:01-02:02:05, 02:05-02:13, 02:15-02:19, 02:21-02:23, 02:25-02:29N	DRB1*10:01:01^w-10:01:04^w, DRB1*10:03^w-10:07^w, DRB1*14:141
	265 bp		*01:02	
15⁵	120 bp	430 bp	*02:03, 03:01:01-03:03	
	180 bp		*02:16	
	210 bp		*02:17	
16	150 bp	430 bp	*01:09, 02:04, 02:19, 02:22	DRB1*03:44
17⁵	120 bp	430 bp	*01:08, 02:06, 02:20	DRB1*03:42, DRB1*03:87, DRB1*13:67, DRB1*14:46
	180 bp		*02:16, 02:23	
18⁵	100 bp	430 bp	*01:09, 02:07, 02:09, 02:21	
19⁷	180 bp	515 bp	*02:08, 02:18, 02:23	DRB1*11:30
	270 bp		*02:14	
20⁷	170 bp	430 bp	*01:01:02:01-01:06, 01:08-01:13, 01:15, 02:09-02:10, 02:16, 03:01:01-03:03	DRB1*03:42, DRB1*03:87, DRB1*11:30, DRB1*13:67, DRB1*14:46
21⁵	120 bp	430 bp	*01:13	
	210 bp		*02:11	
22⁷	185 bp	430 bp	*01:11	

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	245 bp		*01:02, 02:12	
23	195 bp 240 bp	430 bp	*02:13 *01:02	
24	185 bp	430 bp	*02:09, 02:18- 02:19, 02:21, 02:25, 03:01:01- 03:03	DRB1*11:30
25	225 bp 260 bp	430 bp	*02:26 *01:12	
26	235 bp	430 bp	*01:14	DRB1*03:01:02, DRB1*03:02:02, DRB1*03:05:03, DRB1*03:13:02, DRB1*03:17, DRB1*03:24, DRB1*03:27, DRB1*03:35, DRB1*03:81, DRB1*11:27:01, DRB1*11:84:01, DRB1*11:124, DRB1*11:136, DRB1*11:138, DRB1*12:22, DRB1*13:33:02-13:33:03, DRB1*13:61:01, DRB1*13:94:02, DRB1*13:96:02, DRB1*14:38, DRB1*14:47, DRB1*14:50, DRB1*14:98, DRB1*14:127:01
27⁵	90 bp	430 bp	*02:01	
28	235 bp	430 bp	*02:27	DRB1*11:30, DRB1*13:67, DRB1*14:46
29⁸	160 bp	430 bp	*02:15, 02:27	DRB1*10:01:01^w-10:06^w, DRB1*13:106
30	210 bp	430 bp	*02:28	
31⁵	80 bp	430 bp	*02:19, 02:21, 02:25, 02:29N	DRB1*12:34
32⁹	-	-	Negative Control	

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DRB3 SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases. In the presence of a specific amplification the intensity of the control band often decreases.

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³For several DRB1 alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴Due to the sharing of sequence motifs between DRB3 and DRB1 alleles, primer mixes 1 to 5, 7 to 14, 16, 17, 19, 20, 24, 26, 28, 29 and 31 will amplify DRB1 alleles.

⁵HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁶Primer mixes 9, 10 and 13 may have tendencies of unspecific amplifications.

⁷Primer mix 7, 11, 19, 20 and 22 have a tendency to giving rise to primer oligomer formation.

⁸Primer mixes 4 and 29 may give rise to a lower yield of HLA-specific PCR product than the other DRB3 primer mixes.

⁹Primer mix 32 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

‘w’, might be weakly amplified.

‘?’, nucleotide sequence information not available for the primer matching sequence.

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PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	100	125	95	240	95	190	90	120	165	185	270	90
					125		180					145
Length of int. pos. control ¹	515	430	430	430	430	430	515	430	430	515	515	515
5'-primer(s) ²	10(116) 5'-gCT 3'	29(175) 5'-gAT 3'	11(119) 5'-gCg 3'	114(429) 5'-CTg 3'	11(119) 5'-gCg 3'	8(110) 5'-CTC 3'	11(119) 5'-gCg 3'	11(119) 5'-gCg 3'	10(116) 5'-gCT 3'	29(175) 5'-gAC 3'	10(116) 5'-gCT 3'	51(239) 5'-gAg 3'
			11(119) 5'-gTg 3'						10(116) 5'-gCT 3'			
3'-primer(s) ³	29(175) 5'-gTA 3'	57(257) 5'-CgA 3'	28(171) 5'-CTg 3'	181(630) 5'-CTT 3'	28(171) 5'-CTC 3'	57(257) 5'-CgA 3'	27(169) 5'-gTT 3'	37(199) 5'-CAC 3'	51(239) 5'-CCC 3'	77(317) 5'-AgT 3'	86(344) 5'-CCA 3'	66(286) 5'-gAA 3'
	29(175) 5'-gTA 3'	57(257) 5'-CAA 3'	28(171) 5'-ATg 3'		38(202) 5'-gCT 3'		57(257) 5'-CAT 3'			77(318) 5'-TAA 3'		86(344) 5'-CCA 3'
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15	16	17	18	19	20	21	22	23	24
Length of spec. PCR product	270	185	120	150	120	100	180	170	120	185	195	185
		265	180		180		270		210	245	240	
			210									
Length of int. pos. control ¹	515	515	430	430	430	430	515	430	430	430	430	430
5'-primer(s) ²	10(116) 5'-gCT 3'	10(118) 5'-TgT 3'	11(119) 5'-gCT 3'	38(200) 5'-CgC 3'	10(116) 5'-gCT 3'	38(200) 5'-CgC 3'	10(116) 5'-gCT 3'	10(116) 5'-gCT 3'	10(116) 5'-gCT 3'	8(112) 5'-TgC 3'	10(118) 5'-TgT 3'	11(119) 5'-gCT 3'
	10(116) 5'-gCT 3'	38(200) 5'-CgC 3'								10(118) 5'-TgT 3'	25(163) 5'-ggC 3'	
		38(200) 5'-CgC 3'								29(173) 5'-CAT 3'		
3'-primer(s) ³	86(344) 5'-CAC 3'	86(344) 5'-CAC 3'	37(197) 5'-Cgg 3'	74(308) 5'-CCC 3'	36(196) 5'-gTT 3'	57(257) 5'-CgA 3'	55(251) 5'-gCA 3'	51(239) 5'-CCg 3'	36(196) 5'-gAg 3'	77(318) 5'-TAA 3'	77(318) 5'-TAA 3'	58(260) 5'-CCT 3'
			37(197) 5'-CgA 3'		36(196) 5'-gTT 3'		56(256) 5'-gCT 3'	57(257) 5'-CAG 3'	66(286) 5'-gAT 3'			60(266) 5'-Agg 3'
			57(257) 5'-Cag 3'		55(251) 5'-gCA 3'		58(260) 5'-CCT 3'					
			66(286) 5'-gAA 3'		57(257) 5'-CAG 3'		86(344) 5'-CAG 3'					
Well No.	13	14	15	16	17	18	19	20	21	22	23	24

Well No.	25	26	27	28	29	30	31
Length of spec. PCR product	225	235	90	235	160	210	80
	260						
Length of int. pos. control ¹	430	430	430	430	430	430	430
5'-primer(s) ²	10(116) 5'-gCT 3'	12(122) 5'-TAC 3'	163(577) 5'-CAT 3'	10(116) 5'-gCT 3'	38(200) 5'-CgC 3'	123(455) 5'-CTg 3'	51(239) 5'-gAg 3'
						149(534) 5'-Cag 3'	
3'-primer(s) ³	71(299) 5'-gCC 3'	77(317) 5'-AAT 3'	179(624) 5'-ACg 3'	74(308) 5'-CCg 3'	77(317) 5'-Agg 3'	179(624) 5'-ACg 3'	60(266) 5'-Agg 3'
	83(337) 5'-CCg 3'						165(583) 5'-CCg 3'
Well No.	25	26	27	28	29	30	31

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¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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CELL LINE VALIDATION SHEET																					
DRB3 SSP subtyping kit ²																					
					Prod. No.	Well ³															
						1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
						201190801	201080502	201080503	201325532	201080505	201080506	201190807	201080508	201080509	201080510	201080511	201080512	201080513	201441914	201080515	201190816
	IHWC cell line ¹	DRB3																			
1	9001 SA					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280 LK707					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011 E4181324					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275 GU373	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
5	9009 KAS011					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353 SM					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020 QBL	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
8	9025 DEU					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
9	9026 YAR					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107 LKT3					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051 PITOUT					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	9052 DBB					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004 JESTHOM					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071 OLGA					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
15	9075 DKB					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037 SWEIG007	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
17	9282 CTM3953540	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	+	-	-
18	9257 32367	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
19	9038 BM16	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
20	9059 SLE005	*03:01				+	+	-	-	-	-	-	-	-	-	+	-	-	-	+	-
21	9064 AMALA	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
22	9056 KOSE	*02:02	*03:01			+	+	-	+	-	-	-	-	+	+	+	-	+	+	+	-
23	9124 IHL	*02:02				-	-	-	+	-	-	-	-	+	+	-	-	+	+	-	-
24	9035 JBUSH	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
25	9049 IBW9					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
26	9285 WT49	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
27	9191 CH1007					-	-	-	-	-	-	-	-	-	-	-	-	-	W	-	-
28	9320 BEL5GB					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
29	9050 MOU					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
30	9021 RSH	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
31	9019 DUCAF	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
32	9297 HAG	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
33	9098 MT14B					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
34	9104 DHIF	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
35	9302 SSTO					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
36	9024 KT17					-	-	-	-	024	-	-	-	-	-	-	-	-	-	-	-
37	9065 HHKB	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
38	9099 LZL	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
39	9315 CML	*01:01				+	+	+	-	-	-	-	-	-	-	-	-	+	-	-	-
40	9134 WHONP199					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
41	9055 H0301	*03:01				+	+	-	-	-	-	-	-	-	-	+	-	-	-	+	-
42	9066 TAB089					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076 T7526					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057 TEM	*02:24				-	-	-	+	-	-	-	-	+	+	+	+	-	-	-	-
45	9239 SHJO					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013 SCHU					-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045 TUBO	*02:02				-	-	-	-	-	-	-	-	+	+	-	-	+	+	-	-
48	9303 TER-ND					-	-	-	-	9303	-	-	-	-	-	-	-	-	-	-	-

101.121-24/04 – including *Taq* pol., IFU-01
101.121-24u/04u – without *Taq* pol., IFU-02

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[illegible]

¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

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²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

³Cell line TEM, IHW 9057, is profiled as DRB3*02:01 by ECACC ((European Collection of Cell Cultures)). However, Olerup SSP AB has shown that this cell line includes the exon 3 polymorphism corresponding to DRB3*02:24.

Primer mix 4 amplifies DRB1*14 alleles in the KOSE, IHL and TEM cell lines.

The DRB1*10:01 allele is weakly amplified by primer mix 14 in the CH1007 cell line.

No DNAs carrying the alleles to be amplified by primer solutions 5 to 8, 16 to 19, 21 to 23 and 25 to 31 were available. The specificities of the primers in primer solutions 5, 7, 8, 16 to 19, 21, 22, 25, 26, 28, 29 and 31 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solutions 6, 23, 27 and 30 it was only possible to test the 3'-primer, the 5'-primer was not possible to test. One or two of the 3'-primers in primer solutions 1 to 3, 5, 7, 12, 15, 17, 19, 21, 25 and 31 were not possible to test. In primer solutions 2, 3, 14 and 22 one of the 5'-primers was not possible to test. Additional 3'-primers in primer solutions 15, 20 and 24 were tested by separately adding one 5'-primer.

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